# Data sources for the Anopheles-model package

Tom Smith with inputs from Derek Charlwood and Nakul Chitnis

The main source for vector bionomics data is the MAP repository, documented by Massey et al. (1).

## Additional data

Substantial additional data were included in a supplementary database. This includes publications that are not included in the MAP repository, because they did not meet one or other inclusion criterion. These include some very old, and some very recent publications (2-26). A considerable number of additional publications included relate to *An. albimanus* for which there is a substantial literature poorly represented in the MAP repository. The additional data also comprise measures (especially the sac rate, and biting rhythms see below) which were not routinely extracted by Massey et al. (1).

## Post processing

Most of the entomological parameters can be extracted from the format used by MAP, but additional issues arise with:

### Biting rhythms

African data on biting rhythms have recently been compiled by Sherrard-Smith(27). There is no comprehensive database of the biting rhythms for non-African sites, though the MAP files can be searched for publications containing biting rhythms. Biting rhythms were extracted from a convenience sample of publications giving rhythms for a subset of vectors of particular interest (13, 20, 22, 24, 28-33). This is by no means a comprehensive collection of the data that are out there.

### Duration of host-seeking

The duration of host-seeking is estimated from the sac rate where ((using the notation of (34), as described by (4) in (35) and applied also in (24, 35-39). (This ignores the possibility that the mosquitoes might rest for more than 48 hours before oviposition). The duration of host seeking is available only for only a small subset of taxa.

The duration of the gonotrophic cycle is the sum of that of the host-seeking and resting period. Some publications report estimates of the duration of the gonotrophic cycle from either:

1. Mark-recapture(25, 40-48)
2. Cross-correlations in time of biting densities (49)

There are also some publications reporting gonotrophic cycle length where the method used is unclear (50-53). It may be that these estimates really consider only the resting period.

### Duration of resting period

The duration of the resting period can be estimated as follows:

1. Keeping fed mosquitoes to see how long it takes for them to lay eggs (as recommended by WHO(54)). This was carried out by (18, 26, 43, 47, 55-61).
2. The ratio of fed:gravid resting mosquitoes is available for 106 studies in the MAP database for which the fed:gravid ratio is available, corresponding to 30 different Anopheles taxa. Kulkarni et al (2006)(62) and Tchuinkum et al(63) have used the fed:gravid ratio from resting mosquitoes to roughly estimate the duration of the resting period. The Kulkarni et al (2006)(62) analysis gave values of 1:1.4 (low altitude) and 1:4.5 (high altitude) for *An. arabiensis* in Tanzania, translating these into estimates of 2-3 days and 5-6 days.

The logic of the fed:gravid approach is best explained in Tchuinkum et al(63). Assuming negligible mortality while resting and that a whole annual cycle is representatively sampled), the proportion f = fed/(total-unfed), the duration of the cycle should be = 1/f. If we assume daily survival while resting of p<1, and is an integer then:

which is equivalent to:

We can consider the extreme case of assuming all the mortality is at the resting stage, and use this to estimate p from the parous rate, as: . This this leads to an equation for as a function of the parous rate and f:

Feeding in some ‘reasonable’ numbers into this, we get:

|  |  |  |  |
| --- | --- | --- | --- |
|  | M |  |  |
| 0.6 | 0.8 | 1.67 | 1.75 |
| 0.5 | 0.8 | 2.00 | 2.12 |
| 0.4 | 0.8 | 2.50 | 2.68 |
| 0.3 | 0.8 | 3.33 | 3.61 |
| 0.2 | 0.8 | 5.00 | 5.47 |
| 0.1 | 0.8 | 10.00 | 11.05 |
| 0.6 | 0.6 | 1.67 | 1.86 |
| 0.5 | 0.6 | 2.00 | 2.29 |
| 0.4 | 0.6 | 2.50 | 2.93 |
| 0.3 | 0.6 | 3.33 | 4.00 |
| 0.2 | 0.6 | 5.00 | 6.13 |
| 0.1 | 0.6 | 10.00 | 12.51 |

This suggests that could be a reasonable estimate of the duration of the resting period (applicable even if the parous rate has not been determined). The average duration of the full oviposition cycle is then estimated by:

The inputs to the Anopheles package should thus be and (with the constraint that ). Values of the former are obtained either as a temperature-dependent function (various publications have estimated this), or as . Where the sac rate ( is available, can be determined from the above equation. takes real values, but the model is discrete time, with one-day time steps. Effectively this means that we are simulating a Poisson mixture of discrete values (this is broadly supported by published distributions for (58)).

### Estimates of the proportions of mosquitoes resting indoors

The proportions of mosquitoes resting indoors is usually estimated using experimental huts, where exophilic mosquitoes are caught in exit traps. This parameter is relevant for the parameterisation of models of IRS. Publications in the MAP database(1) with exit trap data are (64-67)

## Parameterisation of interventions

Interventions are parameterised by their effects on the input parameter vector:

### Insecticide Treated Nets

The effects of ITNs are parameterised in terms of effectiveness in reducing the availability of humans, , and both pre- and post-prandial killing of mosquitoes and respectively. The effects are based on estimates from experimental hut studies(68). The decay of these effects over time, in terms of attrition, use, physical and chemical integrity is parameterised using the data of PMI net durability studies (7 countries, 8 net types, 23 combinations in all), and also Morgan et al(69)

### Indoor Residual Spraying (IRS)

IRS is also parameterised in terms of reducing the availability of humans, and killing of mosquitoes. 12 different parameterisations (for different insecticide x vector species combinations) are included in the database(70-74).

Following references not sure where they belong:

(75)

(76)

(77)

#### Data used by Swiss TPH in parameterising entomological models

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  |  | Data sources available |  |  |
|  | Swiss TPH publication explaining the paramerisation | Africa | S.E.Asia and Oceania | Americas |
| Mosquito bionomics in the absence of intervention | | | | |
| Human blood index | (78) | (1) | (1) (79, 80), (81) (42) (5, 9) | (1, 19, 73, 82) |
| Biting rhythm | (78) | (27) | (13, 28-32). | (20, 22, 24, 33, 78) |
| Indoor resting | (78) | (1) |  | (20, 22, 23, 83) |
| Parous rate | (78) | (1) | (9, 42, 79) | (25, 84) |
| Duration of Resting period | (78) | (1, 46-49, 55-59, 61-63) | (40, 44, 45, 60) | (18, 25, 26, 43, 47) |
| Duration of host seeking | (34) | (35-38) | (39, 41, 42) | (24) |
| Proportion indoor biting | (78) | (1) | (79-81) |  |
| Models of intervention effects | | | | |
| Effects of LLINs (including PBO nets) on blood feeding and mortality | (85) | (86-98) | (16) | (99) |
| Decay of LLIN effects over time | In prep | PMI studies(attrition, physical and chemical decay for 8 net types in 7 countries); (69) | - | - |
| Availability of hosts to mosquitoes and insecticidal effects of LLINs as functions of insecticide content, holed area, and resistance status | (68, 78) | (68) | - | - |
| IRS effects in reducing human availability and killing of mosquitoes | (78) | (70-74) | - | (78) |

**Information for Palawan vectors is in the xml below (this is the base.xml from Olivier Briet’s Palawan modelling for the Global Fund)**

[<human><component name="**Perfect untreated net**" id="**PUN**"><ITN>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<usage value="**1.0**"/><holeRate sigma="**0**" mean="**0**"/><ripRate sigma="**0**" mean="**0**"/><ripFactor value="**0**"/><initialInsecticide sigma="**0**" mu="**1000**"/><insecticideDecay k="**1**" function="**step**" L="**10000.0**"/><attritionOfNets k="**1**" function="**step**" L="**30d**"/>

<!-- Duration is 0.0822, just over 6/73 = 6\*5=30 days -->

[<anophelesParams propActive="**1.0**" mosquito="**flavirostris**"><twoStageDeterrency>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<entering insecticideScalingFactor="**1**" insecticideFactor="**0.000001**"/>

<!-- Nothing enters -->

<attacking insecticideScalingFactor="**1**" insecticideFactor="**-0.1**" interactionFactor="**0**" holeScalingFactor="**0**" holeFactor="**0**" baseFactor="**0.1**"/></twoStageDeterrency><preprandialKillingEffect insecticideScalingFactor="**0.0**" insecticideFactor="**0**" interactionFactor="**0**" holeScalingFactor="**0**" holeFactor="**0**" baseFactor="**0.0**"/><postprandialKillingEffect insecticideScalingFactor="**0.0**" insecticideFactor="**0**" interactionFactor="**0**" holeScalingFactor="**0**" holeFactor="**0**" baseFactor="**0.0**"/></anophelesParams>[<anophelesParams propActive="**1.0**" mosquito="**balabacensis**"><twoStageDeterrency>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<entering insecticideScalingFactor="**1**" insecticideFactor="**0.000001**"/>

<!-- Nothing enters -->

<attacking insecticideScalingFactor="**1**" insecticideFactor="**-0.1**" interactionFactor="**0**" holeScalingFactor="**0**" holeFactor="**0**" baseFactor="**0.1**"/></twoStageDeterrency><preprandialKillingEffect insecticideScalingFactor="**0.0**" insecticideFactor="**0**" interactionFactor="**0**" holeScalingFactor="**0**" holeFactor="**0**" baseFactor="**0.0**"/><postprandialKillingEffect insecticideScalingFactor="**0.0**" insecticideFactor="**0**" interactionFactor="**0**" holeScalingFactor="**0**" holeFactor="**0**" baseFactor="**0.0**"/></anophelesParams>[<anophelesParams propActive="**1.0**" mosquito="**maculatus**"><twoStageDeterrency>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<entering insecticideScalingFactor="**1**" insecticideFactor="**0.000001**"/>

<!-- Nothing enters -->

<attacking insecticideScalingFactor="**1**" insecticideFactor="**-0.1**" interactionFactor="**0**" holeScalingFactor="**0**" holeFactor="**0**" baseFactor="**0.1**"/></twoStageDeterrency><preprandialKillingEffect insecticideScalingFactor="**0.0**" insecticideFactor="**0**" interactionFactor="**0**" holeScalingFactor="**0**" holeFactor="**0**" baseFactor="**0.0**"/><postprandialKillingEffect insecticideScalingFactor="**0.0**" insecticideFactor="**0**" interactionFactor="**0**" holeScalingFactor="**0**" holeFactor="**0**" baseFactor="**0.0**"/></anophelesParams></ITN></component>[<component name="**LifeNet on An. fluviatilis** " id="**LifeNet**">](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)

<!-- from WHOPES 14th meeting report-->

[<ITN>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<usage value="**0.8**"/><holeRate sigma="**0.7**" mean="**3.25**"/><ripRate sigma="**0.7**" mean="**20**"/><ripFactor value="**1**"/><initialInsecticide sigma="**0.2125**" mu="**8.5**"/>

<!-- WHOPES 14 found a Relative Standard Deviation of 2.5% at a mean of 7.02, thus sigma=0.025\*8.5 for LifeNet-->

<insecticideDecay sigma="**0.8**" mu="**-0.32**" function="**exponential**" L="**2.0**"/>

<!-- Standard value is 1.5 year halflife, but 2 years may be reasonable-->

<attritionOfNets k="**18**" function="**smooth-compact**" L="**18.176**"/>

<!-- Standard value -->

[<anophelesParams propActive="**0.4**" mosquito="**flavirostris**">](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<holeIndexMax value="**145200**"/>[<twoStageDeterrency>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)

<!-- <enteringLogit baseFactor="4.0868" insecticideFactor="-2.8851"/>-->

<entering insecticideScalingFactor="**0.2722518**" insecticideFactor="**0.1553273**"/><attackingLogit insecticideFactor="**1.47636**" interactionFactor="**-0.17521**" holeFactor="**0.47219**" baseFactor="**-0.75405**"/></twoStageDeterrency><preprandialKillingEffectLogit insecticideFactor="**3.44955**" interactionFactor="**-0.08499**" holeFactor="**-0.34248**" baseFactor="**-3.00830**"/><postprandialKillingEffectLogit insecticideFactor="**3.231888**" interactionFactor="**0.008622**" holeFactor="**-0.011861**" baseFactor="**-3.800727**"/></anophelesParams>[<anophelesParams propActive="**0.4**" mosquito="**balabacensis**">](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<holeIndexMax value="**145200**"/>[<twoStageDeterrency>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)

<!-- <enteringLogit baseFactor="4.0868" insecticideFactor="-2.8851"/>-->

<entering insecticideScalingFactor="**0.2722518**" insecticideFactor="**0.1553273**"/><attackingLogit insecticideFactor="**1.47636**" interactionFactor="**-0.17521**" holeFactor="**0.47219**" baseFactor="**-0.75405**"/></twoStageDeterrency><preprandialKillingEffectLogit insecticideFactor="**3.44955**" interactionFactor="**-0.08499**" holeFactor="**-0.34248**" baseFactor="**-3.00830**"/><postprandialKillingEffectLogit insecticideFactor="**3.231888**" interactionFactor="**0.008622**" holeFactor="**-0.011861**" baseFactor="**-3.800727**"/></anophelesParams>[<anophelesParams propActive="**0.4**" mosquito="**maculatus**">](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<holeIndexMax value="**145200**"/>[<twoStageDeterrency>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)

<!-- <enteringLogit baseFactor="4.0868" insecticideFactor="-2.8851"/>-->

<entering insecticideScalingFactor="**0.2722518**" insecticideFactor="**0.1553273**"/><attackingLogit insecticideFactor="**1.47636**" interactionFactor="**-0.17521**" holeFactor="**0.47219**" baseFactor="**-0.75405**"/></twoStageDeterrency><preprandialKillingEffectLogit insecticideFactor="**3.44955**" interactionFactor="**-0.08499**" holeFactor="**-0.34248**" baseFactor="**-3.00830**"/><postprandialKillingEffectLogit insecticideFactor="**3.231888**" interactionFactor="**0.008622**" holeFactor="**-0.011861**" baseFactor="**-3.800727**"/></anophelesParams></ITN></component>[<component id="**PUNcohort**">](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)

<!-- -->

<recruitmentOnly/><subPopRemoval afterYears="**0.075**"/>

<!-- after just more than 25 days (longer is not necessary for TBV effect modeling) -->

</component>[<component id="**ITNcohort**">](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)

<!-- -->

<recruitmentOnly/><subPopRemoval afterYears="**100**"/>

<!-- -->

</component>[<deployment name="**ITNcohort deployment**">](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<component id="**ITNcohort**"/>[<continuous>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<deploy begin="**2016-12-27**" targetAgeYrs="**0.0136986301369863**" coverage="**@ITN\_coverage@**"/></continuous>[<timed>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<deploy time="**2016-12-27**" coverage="**@ITN\_coverage@**" maxAge="**90**" minAge="**0**"/>

<!-- "cohort\_coverage" -->

<!-- The cohort should be recruited one time step earlier than the deployment of the intervention, e.g. 0t -->

</timed></deployment>[<deployment name="**ITN deployment**">](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<component id="**LifeNet**"/>[<timed>](file:///C:\git_repos\vector-models\Intervention_models\LLINparameterisation\Philippines\base.xml)<restrictToSubPop id="**ITNcohort**" complement="**false**"/><deploy repeatEnd="**2035-12-31**" repeatStep="**2y**" time="**@start@**" coverage="**1.0**"/>

<!-- coverage should be 1.0 to have 100% coverage of the cohort subpopulation and timing of the IRS round -->

</timed></deployment>

-<entomology name="P.falciparum cases in Bataraza, Brooke's Point and Rizal in 2015" scaledAnnualEIR="@scaledAnnualEIR@" mode="dynamic">  
<!-- "scaledAnnualEIR" -->

-<vector>  
-<anopheles mosquito="flavirostris" propInfectious="0.000021" propInfected="0.078">  
<!-- Standard values -->

-<seasonality input="EIR" annualEIR="@annualEIR\_flavirostris@">  
<!-- "annualEIR\_flavirostris" -->

-<monthlyValues smoothing="fourier">  
<!-- 2015 P.f. passively detected cases in Bataraza, Brooke's Point and Rizal -->  
<value>  
@s1@</value>  
<value>  
@s2@</value>  
<value>  
@s3@</value>  
<value>  
@s4@</value>  
<value>  
@s5@</value>  
<value>  
@s6@</value>  
<value>  
@s7@</value>  
<value>  
@s8@</value>  
<value>  
@s9@</value>  
<value>  
@s10@</value>  
<value>  
@s11@</value>  
<value>  
@s12@</value>  
</monthlyValues>  
</seasonality>  
-<mosq minInfectedThreshold="0.001">  
<!-- Standard value -->  
<mosqRestDuration value="2"/>  
<!-- Catangui 1971 desrcibes 3 days, but early morning cattle baited trap catches (maybe emptied in the early morning) were counted as 3 days if e.g. collected on the morning of the 8th and eggs were observed on the morning of the 10th, whereas this spans 48 hours. Note that Catangui 1985 Bionomics of malaria vectors in The Philippines states: "The gonotrophic cylce is 48 hours" (!) -->  
<extrinsicIncubationPeriod value="9"/>  
<!-- Clements and Peterson, 1981 used 14d for falciparum in flavirostris, but MOSHKOVSKY (1946)S METHOD Duration (in days) = C / (average daily temperature-B) =111/(28.4-16)= 8.951613-->  
<mosqLaidEggsSameDayProportion value="0.628"/>  
<!-- Catangui 1971. Note that Charlwood et al., 2016 http://malariajournal.biomedcentral.com/articles/10.1186/s12936-016-1389-0 found a value of 0.60 for minimus in Pailin-->  
<mosqSeekingDuration value="0.33"/>  
<!-- Standard value -->  
<mosqSurvivalFeedingCycleProbability value="0.534"/>  
<!-- Torres et al., 1996 parous rate of 0.76 based on 200+7\*200=1600 mosquitoes in Jan-Feb 1993. (0.76/0.72)\*0.43=0.45 in comparison to maculatus. Clements and Peterson 1981 write: "Catangui (1971) analysed the physiological age composition of a population of Anophelesflavirostris at Caloocan, Rizal, in the Philippines, dissecting 1663 females at bait between September and June. Ten percent of females survived 4 gonotrophic cycles, and over 2% survived 6 gonotrophic cycles. Catangui, F. P. (1971). Studies on the gonotrophic cycle of Anopheles minimus flavirostris and the application of physiological age grading technique on the same species. South East Asian Journal of Tropical Medicine and Public Health, 2, 384-392." Thus crudely, 0.10^(1/4)=0.5623413, but they fitted exp(-0.196)^3.5 = 0.5035864. However, the crude data show an average parous rate over a whole year of 0.566, and a fitted value is exp(-0.62810)=0.5336047. This was in a context of DDT spraying. -->  
<availabilityVariance value="0"/>  
<!-- Standard value -->  
<mosqProbBiting mean="0.95" variance="0"/>  
<!-- Standard value -->  
<mosqProbFindRestSite mean="0.95" variance="0"/>  
<!-- Standard value -->  
<mosqProbResting mean="0.99" variance="0"/>  
<!-- Standard value -->  
<mosqProbOvipositing value="0.88"/>  
<!-- Standard value -->  
<mosqHumanBloodIndex value="@mosqHumanBloodIndex\_flavirostris@"/>  
<!-- "mosqHumanBloodIndex.flavirostris" -->  
<!-- Catangui 1985 Bionomics of malaria vectors in The Philippines based on Laurel 1931-1934. Note that Schultz 1993 found a man:carabao ratio of 1:8.19. Thus, with this preference level, with a density ratio of man:carbao of 1.275:1, this yields a HBI of 1.275/(1\*1.275+1\*8.19)= 0.1347 -->  
</mosq>  
-<simpleMPD>  
<developmentDuration value="13"/>  
<!-- Catangui 1985 Bionomics of malaria vectors in The Philippines (average 8-14+48hrs hatching) -->  
<developmentSurvival value="0.467"/>  
<!-- Phasomkulsolsil 2012 value for minimus (related to flavirostris) was 46.7. For sawadwongporni (related to maculatus) on human blood was 79.1%. The value for dirus on human blood was 80.1. Note that cracens (related to dirus) was 87.3 -->  
<femaleEggsLaidByOviposit value="43"/>  
<!-- "43" Salazar, 1989 The malaria situation in The Philippines: a critique (State of the art Malaria) based on Catangui 1985 Bionomics of malaria vectors in The Philippines -->  
<!-- minimum of product of femaleEggsLaidByOviposit and developmentSurvival = 33 !!!-->  
</simpleMPD>  
-<nonHumanHosts name="unprotectedAnimals">  
<mosqRelativeEntoAvailability value="1.0"/>  
<!-- Standard value -->  
<mosqProbBiting value="0.95"/>  
<!-- Standard value -->  
<mosqProbFindRestSite value="0.95"/>  
<!-- Standard value -->  
<mosqProbResting value="0.99"/>  
<!-- Standard value -->  
</nonHumanHosts>  
</anopheles>  
-<anopheles mosquito="maculatus" propInfectious="0.000021" propInfected="0.078">  
<!-- Standard values propInfected="0.078" propInfectious="0.021"-->

-<seasonality input="EIR" annualEIR="@annualEIR\_maculatus@">  
<!-- "annualEIR\_maculatus" -->  
<!-- set to 1 if this is the local species, otherwise to a small number, e.g. 0.001 -->

-<monthlyValues smoothing="fourier">  
<!-- 2015 P.f. passively detected cases in Bataraza, Brooke's Point and Rizal -->  
<value>  
@s1@</value>  
<value>  
@s2@</value>  
<value>  
@s3@</value>  
<value>  
@s4@</value>  
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@s10@</value>  
<value>  
@s11@</value>  
<value>  
@s12@</value>  
</monthlyValues>  
</seasonality>  
-<mosq minInfectedThreshold="0.001">  
<!-- Standard value "0.001" -->  
<mosqRestDuration value="2"/>  
<!-- estimated oviposition cycle was 2.35 plus minus 0.45 days (1.98-3.42 days) from Chiang et al., 1991 -->  
<extrinsicIncubationPeriod value="9"/>  
<!-- Clements and Peterson, 1981 used 14d for falciparum in flavirostris, but MOSHKOVSKYSMETHOD(1946)Duration (in days) = C/(average daily temperature-B) =111/(28.4-16)= 8.951613 -->  
<mosqLaidEggsSameDayProportion value="0.49"/>  
<!-- Charlwood et al., 2016 http://malariajournal.biomedcentral.com/articles/10.1186/s12936-016-1389-0 value for maculatus in Pursat-->  
<mosqSeekingDuration value="0.33"/>  
<!-- Standard value -->  
<mosqSurvivalFeedingCycleProbability value="0.43"/>  
<!-- 0.70 daily survivorship with an estimated oviposition cycle was 2.35 plusminus 0.45 days (1.98-3.42 days) from Chiang et al., 1991 0.7^2.35=0.43. Torres et al., 1996 parous rate of 0.72 based on 50+50\*50=2550 mosquitoes in Jan-Feb 1993 (E.P. Torres et al. : Acta Tropica 63 (1997) 209-220 211: In areas such as Morong and Bataan, residual spraying is done bi-annually with Bendiocarb (Ficam) as the insecticide of choice.) , Charlwood et al., 2016 http://malariajournal.biomedcentral.com/articles/10.1186/s12936-016-1389-0 value of parous rate for maculatus in Pursat was 0.55 on 603 specimens. Shinta and Sukowati 2012 found an average parous rate of 27.9% of maculatus on central Java, presumably in the context of control -->  
<availabilityVariance value="0"/>  
<!-- Standard value -->  
<mosqProbBiting mean="0.95" variance="0"/>  
<!-- Standard value -->  
<mosqProbFindRestSite mean="0.95" variance="0"/>  
<!-- Standard value -->  
<mosqProbResting mean="0.99" variance="0"/>  
<!-- Standard value -->  
<mosqProbOvipositing value="0.88"/>  
<!-- Standard value -->  
<mosqHumanBloodIndex value="@mosqHumanBloodIndex\_maculatus@"/>  
<!-- "mosqHumanBloodIndex\_maculatus" -->  
<!-- Catangui 1985 Bionomics of malaria vectors in The Philippines based on Laurel 1931-1934 Schultz 1993 found a man:carabao preference of maculatus of 1:304.125 with a density ratio of man:carbao of 1.275:1, this yields a HBI of 1.275/(1\*1.275+1\*304.125)= 0.004175 -->  
</mosq>  
-<simpleMPD>  
<developmentDuration value="15"/>  
<!-- Salazar, 1989 The malaria situation in The Philippines: a critique (State of the art Malaria) and Catangui 1985 Bionomics of malaria vectors in The Philippines (average 8-14+48hrs hatching)-->  
<developmentSurvival value="0.791"/>  
<!-- Phasomkulsolsil 2012 value for sawadwongporni (related to maculatus) on human blood was 79.1%. The value for dirus on human blood was 80.1. Note that cracens (related to dirus) was 87.3, minimus (related to flavirostris) was 46.7 -->  
<femaleEggsLaidByOviposit value="43"/>  
<!-- Catangui 1985 Bionomics of malaria vectors in The Philippines -->  
<!-- minimum of product of femaleEggsLaidByOviposit and developmentSurvival = 33 !!! -->  
</simpleMPD>  
-<nonHumanHosts name="unprotectedAnimals">  
<mosqRelativeEntoAvailability value="1.0"/>  
<!-- Standard value -->  
<mosqProbBiting value="0.95"/>  
<!-- Standard value -->  
<mosqProbFindRestSite value="0.95"/>  
<!-- Standard value -->  
<mosqProbResting value="0.99"/>  
<!-- Standard value -->  
</nonHumanHosts>  
</anopheles>  
-<anopheles mosquito="balabacensis" propInfectious="0.000021" propInfected="0.078">  
<!-- Standard values -->

-<seasonality input="EIR" annualEIR="@annualEIR\_balabacensis@">  
<!-- "annualEIR\_balabacensis" -->  
<!-- set to 1 if this is the local species, otherwise 0 -->  
-<monthlyValues smoothing="fourier">  
<!-- 2015 P.f. passively detected cases in Bataraza, Brooke's Point and Rizal -->  
<value>  
@s1@</value>  
<value>  
@s2@</value>  
<value>  
@s3@</value>  
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@s10@</value>  
<value>  
@s11@</value>  
<value>  
@s12@</value>  
</monthlyValues>  
</seasonality>  
-<mosq minInfectedThreshold="0.001">  
<!-- Standard value -->  
<mosqRestDuration value="2"/>  
<!-- Hii et al., 1990: "the oviposition cycle interval 2-3 days" -->  
<extrinsicIncubationPeriod value="9"/>  
<!-- Clements and Peterson, 1981 used 14d for falciparum in flavirostris, but MOSHKOVSKY (1946) S METHOD Duration (in days) = C / (average daily temperature-B) =111/(28.4-16)= 8.951613-->  
<mosqLaidEggsSameDayProportion value="0.34"/>  
<!-- Charlwood et al., 2016 http://malariajournal.biomedcentral.com/articles/10.1186/s12936-016-1389-0 value for dirus in Mondolkiri-->  
<mosqSeekingDuration value="0.33"/>  
<!-- Standard value -->  
<mosqSurvivalFeedingCycleProbability value="0.51"/>  
<!-- Hii et al., 1990 "The estimated survival per oviposition cycle was 0.48-0.54." "which was done in programmatic condition i.e. DDT spraying and surveillance (Hii, personal communication)". Wong 2015 on Sabah found parous rates of 58%, 64% and 65% in village, small plot agriculture and forest settings, respectively, in a context of DDT IRS and LLINs (core prevention in Kudat is ITN and IRS DDT, Hii personal communication) [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4598189/] . Charlwood et al., 2016 http://malariajournal.biomedcentral.com/articles/10.1186/s12936-016-1389-0 parous rate value for dirus in Mondolkiri was 0.58 on 599 specimens-->  
<availabilityVariance value="0"/>  
<!-- Standard value -->  
<mosqProbBiting mean="0.95" variance="0"/>  
<!-- Standard value -->  
<mosqProbFindRestSite mean="0.95" variance="0"/>  
<!-- Standard value -->  
<mosqProbResting mean="0.99" variance="0"/>  
<!-- Standard value -->  
<mosqProbOvipositing value="0.88"/>  
<!-- Standard value -->  
<mosqHumanBloodIndex value="@mosqHumanBloodIndex\_balabacensis@"/>  
<!-- "mosqHumanBloodIndex\_balabacensis" -->  
<!-- Salazar, 1989 The malaria situation in The Philippines: a critique (State of the art Malaria): "balabacensis is highly antropophilic", Schultz 1993 Schultz 1993 found a man:carabao preference of balabacensis of 1:3.3784 with a density ratio of man:carbao of 1.275:1, this yields a HBI of 1.275/(1\*1.275+1\*3.3784)= 0.2739932 -->  
</mosq>  
-<simpleMPD>  
<developmentDuration value="10"/>  
<!-- Phasomkulsolsil 2012 value for dirus on human blood. Note that cracens (related to dirus) was 11d, minimus (related to flavirostris) was 22d and sawadwongporni (related to maculatus) was 15d -->  
<developmentSurvival value="0.71"/>  
<!-- Kanda et al., 1981 for KTD from Kota Belud, Sabah -->  
<!-- Phasomkulsolsil 2012 value for dirus on human blood was 80.1. Note that cracens (related to dirus) was 87.3, minimus (related to flavirostris) was 46.7 and sawadwongporni (related to maculatus) was 79.1% -->  
<femaleEggsLaidByOviposit value="57"/>  
<!-- Kanda et al., 1981 for KTD from Kota Belud, Sabah -->  
<!-- minimum of product of femaleEggsLaidByOviposit and developmentSurvival = 33 !!!-->  
</simpleMPD>  
-<nonHumanHosts name="unprotectedAnimals">  
<mosqRelativeEntoAvailability value="1.0"/>  
<!-- Standard value -->  
<mosqProbBiting value="0.95"/>  
<!-- Standard value -->  
<mosqProbFindRestSite value="0.95"/>  
<!-- Standard value -->  
<mosqProbResting value="0.99"/>

## References

1. Massey NC, Garrod G, Wiebe A, Henry AJ, Huang Z, Moyes CL, et al. A global bionomic database for the dominant vectors of human malaria. Sci Data. 2016;3:160014.

2. Charlwood JD, Nenhep S, Sovannaroth S, Morgan JC, Hemingway J, Chitnis N, et al. 'Nature or nurture': survival rate, oviposition interval, and possible gonotrophic discordance among South East Asian anophelines. Malar J. 2016;15(1):356.

3. Chatterjee S, Chandra G. Role of Anopheles subpictus as a Primary Vector of Malaria in an area in India. Jpn J Trop Med Hyg. 2000;28(3):177-81.

4. Charlwood JD, Birley MH, Dagoro H, Paru R, Holmes PR. Assessing Survival Rates of Anopheles-Farauti (Diptera, Culicidae) from Papua-New-Guinea. Journal of Animal Ecology. 1985;54(3):1003-16.

5. Charlwood JD, Graves PM, Alpers MP. The ecology of the Anopheles punctulatus group of mosquitoes from Papua New Guinea: a review of recent work. PNG Med J. 1986;29(1):19-26.

6. Charlwood JD. Survival rate variation of Anopheles farauti (Diptera: Culicidae) between neighboring villages in coastal Papua New Guinea. J Med Entomol. 1986;23(4):361-5.

7. Dev V, Phookan S, Sharma VP, Anand SP. Physiographic and entomologic risk factors of malaria in Assam, India. Am J Trop Med Hyg. 2004;71(4):451-6.

8. Elyazar IR, Sinka ME, Gething PW, Tarmidzi SN, Surya A, Kusriastuti R, et al. The distribution and bionomics of anopheles malaria vector mosquitoes in Indonesia. Adv Parasitol. 2013;83:173-266.

9. Graves PM, Burkot TR, Saul AJ, Hayes RJ, Carter R. Estimation of Anopheline Survival Rate, Vectorial Capacity and Mosquito Infection Probability from Malaria Vector Infection- Rates in Villages Near Madang, Papua-New-Guinea. Journal Of Applied Ecology. 1990;27(1):134-47.

10. Gunasekaran K, Jambulingam P, Das PK. Distribution of indoor-resting Anopheles fluviatilis in human dwellings and its implication on indoor residual spray. Indian J Malariol. 1995;32(1):42-6.

11. Gunathilaka N, Denipitiya T, Hapugoda M, Abeyewickreme W, Wickremasinghe R. Determination of the foraging behaviour and blood meal source of malaria vector mosquitoes in Trincomalee District of Sri Lanka using a multiplex real time polymerase chain reaction assay. Malar J. 2016;15:242.

12. Ismail IAH, Notananda V, Schapens J. Studies of Malaria and Responses of Anopheles balabacensis and Anopheles minimus to DDT residual spraying in Thailand. Geneva; 1973 1973. Report No.: WHO/MAL/73.810.

13. Singh N, Mishra AK, Chand SK, Sharma VP. Population dynamics of Anopheles culicifacies and malaria in the tribal area of central India. J Am Mosq Control Assoc. 1999;15(3):283-90.

14. Mahmood F, Reisen WK. Duration of the gonotrophic cycles of Anopheles culifacies Giles and Anopheles stephensi Liston, with observations on reproductive activity and survivorship during winter in Punjab Province, Pakistan. Mosquito News. 1981;41(1):41-50.

15. Ndoen E, Wild C, Dale P, Sipe N, Dale M. Mosquito Longevity, Vector Capacity, and Malaria Incidence in West Timor and Central Java, Indonesia. ISRN Public Health. 2012;2012(Article ID 143863):1-5.

16. Van Bortel W, Chinh VD, Berkvens D, Speybroeck N, Trung HD, Coosemans M. Impact of insecticide-treated nets on wild pyrethroid resistant Anopheles epiroticus population from southern Vietnam tested in experimental huts. Malar J. 2009;8:248.

17. WHARTON RH. The habits of adult mosquitoes in Malaya. I. Observations on anophelines in window-trap huts and at cattle-sheds. Ann Trop Med Parasitol. 1951;45(2):141-54.

18. Mekuria Y, Granados R, Tidwell MA, Williams DC, Wirtz RA, Roberts DR. Malaria transmission potential by Anopheles mosquitoes of Dajabon, Dominican Republic. J Am Mosq Control Assoc. 1991;7(3):456-61.

19. Ricciardi ID. Definicion de los habitos alimentares de anophelinos de Guatemala y Republica Dominicana, por tecnicas de gel-precipitation. Rev Microbiol. 1971;2(3):107-12.

20. Hobbs JH, Sexton JD, St JY, Jacques JR. The biting and resting behavior of Anopheles albimanus in northern Haiti. J Am Mosq Control Assoc. 1986;2(2):150-3.

21. Muirhead-Thomson RC, Mercier EC. Factors in malaria transmission by Anopheles albimanus in Jamaica. II. Ann Trop Med Parasitol. 1952;46(3):201-13.

22. Taylor RT. The ecology of Anopheles albimanus (Wied.) in Haiti. Mosquito News. 1966;26(3):393-7.

23. Bown DN, Rodriguez MH, Arredondo-Jimenez JI, Loyola EG, Rodriguez MC. Intradomiciliary behavior of Anopheles albimanus on the coastal plain of southern Mexico: implications for malaria control. J Am Mosq Control Assoc. 1993;9(3):321-4.

24. Molez JF, Desenfant P, Jacques JR. Bio-ecology of Anopheles albimanus Wiedeman, 1820 (Diptera : Culicidae) in Haiti (Hispaniola). Bulletin de la Societe de Pathologie Exotique. 1998;91(4):334-9.

25. Rodriguez MH, Bown DN, Arredondo-Jimenez JI, Villarreal C, Loyola EG, Frederickson CE. Gonotrophic cycle and survivorship of Anopheles albimanus (Diptera: Culicidae) in southern Mexico. J Med Entomol. 1992;29(3):395-9.

26. Rua GL, Quinones ML, Velez ID, Zuluaga JS, Rojas W, Poveda G, et al. Laboratory estimation of the effects of increasing temperatures on the duration of gonotrophic cycle of Anopheles albimanus (Diptera: Culicidae). Mem Inst Oswaldo Cruz. 2005;100(5):515-20.

27. Sherrard-Smith E, Skarp JE, Beale AD, Fornadel C, Norris LC, Moore SJ, et al. Mosquito feeding behavior and how it influences residual malaria transmission across Africa. Proc Natl Acad Sci U S A. 2019.

28. Ritthison W, Tainchum K, Manguin S, Bangs MJ, Chareonviriyaphap T. Biting patterns and host preference of Anopheles epiroticus in Chang Island, Trat Province, eastern Thailand. J Vector Ecol. 2014;39(2):361-71.

29. Ndoen E, Wild C, Dale P, Sipe N, Dale M. Dusk to dawn activity patterns of anopheline mosquitoes in West Timor and Java, Indonesia. Southeast Asian J Trop Med Public Health. 2011;42(3):550-61.

30. Dev V. Anopheles minimus: Its bionomics and role in the transmission of malaria in Assam, India. Bulletin of the World Health Organization. 1996;74(1):61-6.

31. Manh CD, Beebe NW, Van VN, Quang TL, Lein CT, Nguyen DV, et al. Vectors and malaria transmission in deforested, rural communities in north-central Vietnam. Malar J. 2010;9:259.

32. Hii JL, Smith T, Mai A, Ibam E, Alpers MP. Comparison between anopheline mosquitoes (Diptera: Culicidae) caught using different methods in a malaria endemic area of Papua New Guinea. Bull Entomol Res. 2000;90(3):211-9.

33. Desenfant P. Rôle et bioécologie de *A. albimanus* (Wiedemann, 1820) vecteur du paludisme en Haiti: Université de Paris-Sud; 1988.

34. Chitnis N, Smith T, Steketee R. A mathematical model for the dynamics of malaria in mosquitoes feeding on a heterogeneous host population. J Biol Dyn. 2008;2(3):259-85.

35. Charlwood JD, Smith T, Billingsley P, Takken W, Lyimo E, Meuwissen J. Survival and infection probabilities of anthropophagic anophelines from an area of high prevalence of *Plasmodium falciparum* in humans. Bull Entomol Res. 1997;87:445-53.

36. Charlwood JD, Kihonda J, Sama S, Billingsley PF, Hadji H, Verhave JP, et al. The Rise and Fall of Anopheles Arabiensis (Diptera, Culicidae) in A Tanzanian Village. Bulletin of Entomological Research. 1995;85(1):37-44.

37. Charlwood JD, Pinto J, Sousa CA, Ferreira C, Gil V, do Rosario VE. Mating does not affect the biting behaviour of Anopheles gambiae from the islands of Sao Tome and Principe, West Africa. Annals of Tropical Medicine and Parasitology. 2003;97(7):751-6.

38. Charlwood JD, Pinto J, Sousa CA, Ferreira C, Petrarca V, Rosario VD. 'A mate or a meal' - Pre-gravid behaviour of female Anopheles gambiae from the islands of Sao Tome and Principe, West Africa. Malaria Journal. 2003;2.

39. Charlwood JD, Nenhep S, Sovannaroth S, Morgan JC, Hemingway J, Chitnis N, et al. 'Nature or nurture': survival rate, oviposition interval, and possible gonotrophic discordance among South East Asian anophelines. Malaria Journal. 2016;15.

40. Birley MH, Charlwood JD. The Effect of Moonlight and Other Factors on the Oviposition Cycle of Malaria Vectors in Madang, Papua-New-Guinea. Annals of Tropical Medicine and Parasitology. 1989;83(4):415-22.

41. Charlwood JD, Graves PM, Birley MH. Capture-Recapture Studies with Mosquitos of the Group of Anopheles-Punctulatus Donitz (Diptera, Culicidae) from Papua- New-Guinea. Bulletin of Entomological Research. 1986;76(2):211-27.

42. Charlwood JD, Graves PM. The effect of permethrin-impregnated bednets on a population of Anopheles farauti in coastal Papua New Guinea. Med Vet Entomol. 1987;1(3):319-27.

43. Fernandez-Salas I, Rodriguez MH, Roberts DR. Gonotrophic cycle and survivorship of Anopheles pseudopunctipennis (Diptera: Culicidae) in the Tapachula foothills of southern Mexico. J Med Entomol. 1994;31(3):340-7.

44. Chiang GL, Loong KP, Chan ST, Eng KL, Yap HH. Capture-recapture studies with Anopheles maculatus Theobald (Diptera: Culicidae) the vector of malaria in peninsular Malaysia. Southeast Asian J Trop Med Public Health. 1991;22(4):643-7.

45. Jaal Z, MacDonald WW. A mark-release-recapture experiment with Anopheles lesteri paraliae in northwest Peninsular Malaysia. Ann Trop Med Parasitol. 1992;86(4):419-24.

46. Quinones ML, Lines JD, Thomson MC, Jawara M, Morris J, Greenwood BM. Anopheles gambiae gonotrophic cycle duration, biting and exiting behaviour unaffected by permethrin-impregnated bednets in The Gambia. Med Vet Entomol. 1997;11(1):71-8.

47. Santos RL, Forattini OP, Burattini MN. Laboratory and field observations on duration of gonotrophic cycle of Anopheles albitarsis s.l. (Diptera: Culicidae) in southeastern Brazil. J Med Entomol. 2002;39(6):926-30.

48. Toure YT, Dolo G, Petrarca V, Traore SF, Bouare M, Dao A, et al. Mark-release-recapture experiments with Anopheles gambiae s.l. in Banambani Village, Mali, to determine population size and structure. Med Vet Entomol. 1998;12(1):74-83.

49. Bockarie M, Service MW, Barnish G, Toure Y. Vectorial capacity and entomological inoculation rates of Anopheles gambiae in a high rainfall forested area of southern Sierra Leone. Trop Med Parasitol. 1995;46(3):164-71.

50. Ijumba JN, Mwangi RW, Beier JC. Malaria Transmission Potential of Anopheles Mosquitos in the Mwea-Tebere Irrigation Scheme, Kenya. Medical and Veterinary Entomology. 1990;4(4):425-32.

51. Ameneshewa B, Service MW. Blood-feeding behaviour of Anopheles arabiensis Patton (Diptera: Culicidae) in central Ethiopia. Journal of African Zoology. 1997;111(3):235-45.

52. Hii JL, Birley MH, Kanai L, Foligeli A, Wagner J. Comparative effects of permethrin-impregnated bednets and DDT house spraying on survival rates and oviposition interval of Anopheles farauti No. 1 (Diptera:Culicidae) in Solomon Islands. Ann Trop Med Parasitol. 1995;89(5):521-9.

53. Chandra G. Age composition of incriminated malaria vector in a rural foothills in West Bengal, India. Indian Journal of Medical Research. 2008;127(6):607-9.

54. Organisation WH. Manual on Practical Entomology in Malaria. Part II: Methods and Techniques. 1975.

55. Manga L, Toto JC, Le Goff G, Brunhes J. The bionomics of Anopheles funestus and its role in malaria transmission in a forested area of southern Cameroon. Trans R Soc Trop Med & Hyg. 1997;91(4):387-8.

56. Ijumba JN, Mosha FW, Lindsay SW. Malaria transmission risk variations derived from different agricultural practices in an irrigated area of northern Tanzania. Med Vet Entomol. 2002;16(1):28-38.

57. Afrane YA, Lawson BW, Githeko AK, Yan GY. Effects of microclimatic changes caused by land use and land cover on duration of gonotrophic cycles of Anopheles gambiae (Diptera : culicidae) in western Kenya highlands. Journal of Medical Entomology. 2005;42(6):974-80.

58. Beier JC. Frequent blood-feeding and restrictive sugar-feeding behavior enhance the malaria vector potential of Anopheles gambiae s.l. and An. funestus (Diptera:Culicidae) in western Kenya. J Med Entomol. 1996;33(4):613-8.

59. Mendis C, Jacobsen JL, Gamage-Mendis A, Bule E, Dgedge M, Thompson R, et al. Anopheles arabiensis and An. funestus are equally important vectors of malaria in Matola coastal suburb of Maputo, southern Mozambique. Med Vet Entomol. 2000;14(2):171-80.

60. Ree HI, Hwang UW, Lee IY, Kim TE. Daily survival and human blood index of Anopheles sinensis, the vector species of malaria in Korea. J Am Mosq Control Assoc. 2001;17(1):67-72.

61. Tanga MC, Ngundu WI, Tchouassi PD. Daily survival and human blood index of major malaria vectors associated with oil palm cultivation in Cameroon and their role in malaria transmission. Trop Med Int Health. 2011;16(4):447-57.

62. Kulkarni MA, Kweka E, Nyale E, Lyatuu E, Mosha FW, Chandramohan D, et al. Entomological evalution of malaria vectors at different altitudes in Hal District, Northeastern Tanzania. Journal of Medical Entomology. 2006;43(3):580-8.

63. Tchuinkam T, Simard F, Lele-Defo E, Tene-Fossog B, Tateng-Ngouateu A, Antonio-Nkondjio C, et al. Bionomics of Anopheline species and malaria transmission dynamics along an altitudinal transect in Western Cameroon. BMC Infect Dis. 2010;10:119.

64. Mnzava AE, Rwegoshora RT, Wilkes TJ, Tanner M, Curtis CF. Anopheles arabiensis and An. gambiae chromosomal inversion polymorphism, feeding and resting behaviour in relation to insecticide house-spraying in Tanzania. Med Vet Entomol. 1995;9(3):316-24.

65. Sreehari U, Razdan RK, Mittal PK, Ansari MA, Rizvi MM, Dash AP. Impact of Olyset nets on malaria transmission in India. J Vector Borne Dis. 2007;44(2):137-44.

66. Akogbeto M, Padonou GG, Bankole HS, Gazard DK, Gbedjissi GL. Dramatic decrease in malaria transmission after large-scale indoor residual spraying with bendiocarb in Benin, an area of high resistance of Anopheles gambiae to pyrethroids. Am J Trop Med Hyg. 2011;85(4):586-93.

67. Sachs P, Diaz Rodriguez GA, Briceno I, King R, Achee NL, Grieco JP. Comparison of experimental hut entrance and exit behavior between Anopheles darlingi from the Cayo District, Belize, and Zungarococha, Peru. J Am Mosq Control Assoc. 2013;29(4):319-27.

68. Randriamaherijaona S, Briet OJ, Boyer S, Bouraima A, N'Guessan R, Rogier C, et al. Do holes in long-lasting insecticidal nets compromise their efficacy against pyrethroid resistant Anopheles gambiae and Culex quinquefasciatus? Results from a release-recapture study in experimental huts. Malar J. 2015;14:332.

69. Morgan J, Abilio AP, do Rosario Pondja M, Marrenjo D, Luciano J, Fernandes G, et al. Physical durability of two types of long-lasting insecticidal nets (LLINs) three years after a mass LLIN distribution campaign in Mozambique, 2008-2011. Am J Trop Med Hyg. 2015;92(2):286-93.

70. Bown DN, Rodriguez MH, Arredondo-Jimenez JI, Loyola EG, Rodriguez MC. Age structure and abundance levels in the entomological evaluation of an insecticide used in the control of Anopheles albimanus in southern Mexico. J Am Mosq Control Assoc. 1991;7(2):180-7.

71. Agossa FR, Aikpon R, Azondekon R, Govoetchan R, Padonnou GG, Oussou O, et al. Efficacy of various insecticides recommended for indoor residual spraying: pirimiphos methyl, potential alternative to bendiocarb for pyrethroid resistance management in Benin, West Africa. Transactions of the Royal Society of Tropical Medicine and Hygiene. 2014;108(2):84-91.

72. Tchicaya E, Nsanzabana C, Smith TA, Donze J, de Hipsl ML, Muller P, et al. Micro-encapsulated pirimiphos-methyl shows high insecticidal efficacy against pyrethroid-resistant malaria vectors. Tropical Medicine & International Health. 2015;20:20-.

73. Bangs MJ. The susceptibility and behavioral response of *Anopheles albimanus* Weidemann and *Anopheles vestitipennis* Dyar and Knab (Diptera: Culicidae) to insecticides in northern Belize, Central America. Ann Arbor: University of Michigan; 1999.

74. Kuhlow F. Field experiments on the behavior of malaria vectors in an unsprayed hut and in a hut sprayed with DDT in Northern Nigeria. Bull World Health Organ. 1962;26:93-102.

75. Charlwood JD, Dagoro H, Paru R. Blood-Feeding and Resting Behavior in the Anopheles-Punctulatus Donitz Complex (Diptera, Cullicidae) from Coastal Papua-New- Guinea. Bulletin of Entomological Research. 1985;75(3):463-75.

76. Bockarie MJ, Alexander N, Bockarie F, Ibam E, Barnish G, Alpers M. The late biting habit of parous Anopheles mosquitoes and pre-bedtime exposure of humans to infective female mosquitoes. Trans R Soc Trop Med Hyg. 1996;90(1):23-5.

77. Cooper RD, Frances SP. Malaria vectors on Buka and Bougainville Islands, Papua New Guinea. J Am Mosq Control Assoc. 2002;18(2):100-6.

78. Briet OJT, Impoinvil DE, Chitnis N, Pothin E, Lemoine JF, Frederic J, et al. Models of effectiveness of interventions against malaria transmitted by Anopheles albimanus. Malar J. 2019;18(1):263.

79. Bugoro H, Cooper RD, Butafa C, Iro'ofa C, Mackenzie DO, Chen CC, et al. Bionomics of the malaria vector Anopheles farauti in Temotu Province, Solomon Islands: issues for malaria elimination. Malar J. 2011;10:133.

80. Bugoro H, Hii JL, Butafa C, Iro'ofa C, Apairamo A, Cooper RD, et al. The bionomics of the malaria vector Anopheles farauti in Northern Guadalcanal, Solomon Islands: issues for successful vector control. Malar J. 2014;13:56.

81. Hii JL, Smith T, Mai A, Mellor S, Lewis D, Alexander N, et al. Spatial and temporal variation in abundance of Anopheles (Diptera:Culicidae) in a malaria endemic area in Papua New Guinea. J Med Entomol. 1997;34(2):193-205.

82. Grieco JP, Achee NL, Andre RG, Roberts DR. Host feeding preferences of Anopheles species collected by manual aspiration, mechanical aspiration, and from a vehicle-mounted trap in the Toledo District, Belize, Central America. J Am Mosq Control Assoc. 2002;18(4):307-15.

83. Muirhead-Thomson RC, Mercier EC. Factors in malaria transmission by Anopheles albimanus in Jamaica. Part I. Ann Trop Med Parasitol. 1952;46(2):103-16.

84. Mekuria Y, Tidwell MA, Williams DC, Mandeville JD. Bionomic studies of the Anopheles mosquitoes of Dajabon, Dominican Republic. J Am Mosq Control Assoc. 1990;6(4):651-7.

85. Briet OJ, Penny MA, Hardy D, Awolola TS, Van BW, Corbel V, et al. Effects of pyrethroid resistance on the cost effectiveness of a mass distribution of long-lasting insecticidal nets: a modelling study. Malar J. 2013;12:77.

86. Tami A, Mubyazi G, Talbert A, Mshinda H, Duchon S, Lengeler C. Evaluation of Olyset insecticide-treated nets distributed seven years previously in Tanzania. Malar J. 2004;3:19.:19.

87. Kilian A, Byamukama W, Pigeon O, Gimnig J, Atieli F, Koekemoer LL, et al. Evidence for a useful life of more than three years for a polyester-based long-lasting insecticidal mosquito net in Western Uganda. Malar J. 2011;10(1):299.

88. Kilian A, Byamukama W, Pigeon O, Atieli F, Duchon S, Phan C. Long-term field performance of a polyester-based long-lasting insecticidal mosquito net in rural Uganda. Malar J. 2008;7:49.:49.

89. Corbel V, Chabi J, Dabire RK, Etang J, Nwane P, Pigeon O, et al. Field efficacy of a new mosaic long-lasting mosquito net (PermaNet 3.0) against pyrethroid-resistant malaria vectors: a multi centre study in Western and Central Africa. Malar J. 2010;9:113.:113.

90. Irish S, N'Guessan R, Boko P, Metonnou C, Odjo A, Akogbeto M, et al. Loss of protection with insecticide-treated nets against pyrethroid-resistant *Culex quinquefasciatus* mosquitoes once nets become holed: an experimental hut study. Parasit Vectors. 2008;1(1):17.

91. N'Guessan R, Asidi A, Boko P, Odjo A, Akogbeto M, Pigeon O, et al. An experimental hut evaluation of PermaNet((R)) 3.0, a deltamethrin-piperonyl butoxide combination net, against pyrethroid-resistant *Anopheles gambiae* and *Culex quinquefasciatus* mosquitoes in southern Benin. Trans R Soc Trop Med Hyg. 2010;104(12):758-65.

92. Tungu P, Magesa S, Maxwell C, Malima R, Masue D, Sudi W, et al. Evaluation of PermaNet 3.0 a deltamethrin-PBO combination net against *Anopheles gambiae* and pyrethroid resistant *Culex quinquefasciatus* mosquitoes: an experimental hut trial in Tanzania. Malar J. 2010;9:21.:21.

93. Kitau J, Oxborough RM, Tungu PK, Matowo J, Malima RC, Magesa SM, et al. Species Shifts in the Anopheles gambiae Complex: Do LLINs Successfully Control Anopheles arabiensis? Plos One. 2012;7(3).

94. Adeogun AO, Olojede JB, Oduola AO, Awolola TS. Efficacy of a combination long lasting insecticidal net (PermaNet® 3.0) against pyrethroid resistant Anopheles gambiae s.s. and Culex quinquefasciatus: an experimental hut trial in Nigeria. . Nigerian Journal of Clinical & Biomedical Research. 2012;6:37-50.

95. Koudou BG, Koffi AA, Malone D, Hemingway J. Efficacy of PermaNet(R) 2.0 and PermaNet(R) 3.0 against insecticide-resistant *Anopheles gambiae* in experimental huts in Cote d'Ivoire. Malar J. 2011;10:172.:172.

96. Chandre F, Darrier F, Manga L, Akogbeto M, Faye O, Mouchet J, et al. Status of pyrethroid resistance in *Anopheles gambiae sensu lato*. Bull World Health Organ. 1999;77(3):230-4.

97. Lines JD, Curtis CF, Myamba J, Njau R. Tests of repellent or insecticide impregnated curtains, bednets and anklets against malaria vectors in Tanzania. 1985 1985. Report No.: WHO/VBC/85.920.

98. Lines JD, Myamba J, Curtis CF. Experimental hut trials of permethrin-impregnated mosquito nets and eave curtains against malaria vectors in Tanzania. Med Vet Entomol. 1987;1(1):37-51.

99. Arredondo-Jimenez JI, Rodriguez MH, Loyola EG, Bown DN. Behaviour of Anopheles albimanus in relation to pyrethroid-treated bednets. Med Vet Entomol. 1997;11(1):87-94.